

# Liquid Chromatography

# Introduction

Chromatography is an important group of methods to separate closely related components of complex mixtures. The following example simulates the separation of species in High Performance Liquid Chromatography (HPLC). In this technique an injector introduces a sample as a zone in a liquid mobile phase. The mobile phase containing the sample zone is pumped through a column containing a solid stationary phase; Figure 1 shows a diagram of such an instrument.



# Figure 1: Diagram of an HPLC system.

The mobile and stationary phases are chosen so that the samples are distributed to varying degrees between the two phases. Those components that strongly adsorb to the stationary phase move only slowly with the flow of the mobile phase, and those that are weakly adsorbed move more rapidly. As the sample zones progress through the column, the components are separated into discrete zones that are recognized by a detector, situated beyond the outlet of the column.

# Model Definition

This model studies the separation of two species under conditions of nonlinear chromatography in a 1D geometry. The Transport of Diluted Species in Porous Media interface is used, with convection and adsorption in porous media accounted for.

The equation for analyte transport through a chromatographic column, with constant porosity, is computed by:

$$(\varepsilon + \rho k_{\mathrm{P},i})\frac{\partial c_i}{\partial t} + \mathbf{u} \cdot \nabla c_i = \nabla \cdot \left[ \left( D_{\mathrm{D},i} + \frac{\varepsilon}{\tau_{\mathrm{F},i}} D_{\mathrm{F},i} \right) \nabla c_i \right] + R_i + S_i$$
(1)

Here,  $c_i$  is the concentration of component *i* (SI unit: mol/m<sup>3</sup>),  $\varepsilon$  is the porosity,  $\rho$  is the density of the media within the column (for the mix of liquid and solid matrix, SI unit: kg/m<sup>3</sup>),  $k_{P,i}$  is an adsorption isotherm, and **u** is the volume average velocity of the fluid phase (SI unit: m/s). The second term on the right-hand side describes the mixing of the

solutes, including mechanical mixing (dispersion) and molecular diffusion. The two last terms on the right-hand side are a reaction rate term and a fluid source term.

Equation 1 comes from the following derivation:

First, consider the dispersion of the chromatographic zone to be negligible as it progresses through the column. The mass transport equation will then take the form:

$$S\frac{\rho_p(1-\varepsilon)}{\varepsilon} \cdot \frac{\partial n_i}{\partial t} + \frac{\partial c_i}{\partial t} = -\frac{v}{\varepsilon A} \cdot \frac{\partial c_i}{\partial x}$$

where *S* denotes the specific surface area of the particles in the column (SI unit:  $m^2/kg$ ),  $\rho_p$  denotes the density of the solid particles (SI unit:  $kg/m^3$ ),  $\varepsilon$  equals the column porosity, *A* gives the inner area of the column tube,  $n_i$  equals the analyte concentration in the stationary phase of component *i* (SI unit:  $mol/m^2$ ), *v* describes the mobile phase flow (SI unit:  $m^3/s$ ), and  $c_i$  equals the analyte concentration in the mobile phase of component *i* (SI unit:  $mol/m^3$ ).

Equation 1 is defined according to the ideal model for chromatography that assumes that the equilibrium for the analyte between the mobile and stationary phases is immediate, that is:

$$\frac{\partial c_{\mathrm{P},i}}{\partial t} = \frac{\partial c_{\mathrm{P},i}}{\partial c_{i}} \cdot \frac{\partial c_{i}}{\partial t} = k_{\mathrm{P},i} \left( \frac{\partial c_{i}}{\partial t} \right)$$

where  $c_{P,i}$  is the concentration of the component adsorbed to the solid (moles per dry unit).

The mass transport equation for the ideal chromatography model therefore becomes:

$$\left(1 + S\frac{\rho_p(1-\varepsilon)}{\varepsilon} \cdot \frac{dn}{dc}\right) \cdot \frac{\partial c_i}{\partial t} = -\frac{v}{\varepsilon A}\frac{\partial c_i}{\partial x}$$

The dispersion or band broadening of the analyte zone is a result of a great number of random processes that the analyte experiences (for example, inhomogeneous flow and diffusion in pores and the mobile phase). It is therefore possible to formally express the band broadening as a diffusion process with an effective diffusion constant,  $D_{\rm eff}$ . Thus,  $D_{\rm eff}$  is a measure of the chromatographic system's efficiency for a particular analyte. This constant is closely related to the concept of the height equivalent of a theoretical plate, H, that is customarily used in chromatographic practice. It can be shown that:

$$D_{\rm eff} = \frac{H v_{zi}}{2}$$

where  $v_{zi}$  is the migration velocity of the analyte zone through the column. A mass balance that includes the zone-dispersion term gives the following equation:

$$\left(1 + \Phi \cdot \frac{dn}{dc}\right) \cdot \frac{\partial c_i}{\partial t} = -v_1 \frac{\partial c_i}{\partial x} + D_{\text{eff}} \frac{\partial^2 c_i}{\partial x^2}$$

Here  $\Phi = S\rho(1-\epsilon)/\epsilon$  denotes the phase ratio of the column (SI unit: m<sup>2</sup>/m<sup>3</sup>),  $v_1 = v/(\epsilon A)$  gives the linear velocity of the mobile phase in the column (SI unit: m/s), and  $D_{\text{eff}}$  is the effective diffusion constant (SI unit: m<sup>2</sup>/s).

This first example covers two components. The adsorption isotherm for both components is assumed to follow a Langmuir adsorption isotherm, that is,

$$n_i = \frac{n_{0i}K_ic_i}{1 + K_ic_i}$$

and

$$\frac{dn_i}{dc_i} = \frac{n_{0i}K_i}{\left(1 + K_ic_i\right)^2}$$

where  $K_i$  is the adsorption constant for component *i* (SI unit: m<sup>3</sup>/mol), and  $n_{0i}$  is the monolayer capacity of the stationary phase for component *i* (SI unit: mol/m<sup>2</sup>).

Using an effective zone-dispersion term and the  $k_{P,i}$  adsorption isotherm notation gives Equation 1 without the reaction rate and a fluid source terms:

$$(\varepsilon + \rho k_{\mathrm{P},i})\frac{\partial c_i}{\partial t} + u\frac{\partial c_i}{\partial x} = \frac{\partial}{\partial x} \left( D_{\mathrm{eff},i}\frac{\partial c_i}{\partial x} \right)$$

# INPUT DATA

TABLE I. INPUT DATA

This example looks at the progress of the chromatographic zone within the column. The physical data for the column correspond to a 12 cm-by-4 mm inner diameter column filled with 5  $\mu$ m porous particles. The rest of the input data appear in Table 1.

NAME	VALUE	
S	100 m <sup>2</sup> /g	
$ ho_{ m p}$	2300 kg/m <sup>3</sup>	
ε	0.6	
$v_1$	1.322 mm/s	

#### 4 | LIQUID CHROMATOGRAPHY

TABLE I: INPUT DATA.

NAME	VALUE
$D_{\mathrm{eff1}}$	I ⋅ 10 <sup>-8</sup> m <sup>2</sup> /s
$D_{ m eff2}$	I · 10 <sup>-8</sup> m <sup>2</sup> /s
$K_1$	0.04 m <sup>3</sup> /mol
$K_2$	0.05 m <sup>3</sup> /mol
<i>n</i> <sub>01</sub>	I ⋅ I 0 <sup>-6</sup> mol/m <sup>2</sup>
$n_{02}$	5·10 <sup>-7</sup> mol/m <sup>2</sup>

The injector concentrations for the two components are described by a normal distribution and are set up with the help of a Gaussian pulse function with an amplitude of 1 (Figure 2).



Figure 2: Injection pulse with amplitude 1.

The maximum inlet concentrations are  $1 \text{ mol/m}^3$  and  $10 \text{ mol/m}^3$  for components 1 and 2, respectively.

Figure 3 shows the mobile concentration zones of component 1 at various times. Initially, the concentration is zero in the column and at approximately 5 s the whole component mixture has been injected through the leftmost boundary (the inlet). At approximatively 385 s, the first trace of component 1 exits the rightmost boundary (the outlet). The zones are nearly symmetrical and normally distributed, indicating that the solution is affected by an almost linear adsorption isotherm.



Figure 3: The concentration of component 1 in the mobile phase at various times.

In Figure 4, the zones for both of the components are displayed at various times. For the present conditions a clear separation of the two components occurs within the column; the component zones do not overlap as they reach the outlet. As an example, at t=300 s, component 2 is no longer present in the mobile phase of the column, while most of component 1 still remains. Here, it is shown that component 2 depends on a nonlinear



adsorption isotherm, quickly obtaining an asymmetrical mobile concentration zone (cf. Figure 2).

Figure 4: The concentrations of components 1 (solid) and 2 (dashed) in the mobile phase at various times.

The concentrations that are registered over time by the detector unit in the HPLC system are displayed in Figure 5. The first peak is that of component 1 and the second that of component 2.



Figure 5: Concentration at the outlet of the column, that is, the concentration monitored by the system detector.

# References

1. D. DeVault, "The Theory of Chromatography," J. Am. Chemical Soc., vol. 65, pp. 532–540, 1943.

2. S. Golshan-Shirazi and G. Guiochon, "Analytical solution for the ideal model of chromatography in the case of a Langmuir isotherm," *Analytical Chemistry*, vol. 60, no. 21, pp. 2364–2374, 1988.

3. B. Lin and G. Guiochon, *Modeling for Preparative Chromatography*, Elsevier Publishing, Amsterdam, 2003.

Application Library path: Chemical\_Reaction\_Engineering\_Module/ Mixing\_and\_Separation/liquid\_chromatography\_1

# Modeling Instructions

From the File menu, choose New.

## NEW

In the New window, click 🔗 Model Wizard.

# MODEL WIZARD

- I In the Model Wizard window, click ID.
- 2 In the Select Physics tree, select Chemical Species Transport> Transport of Diluted Species in Porous Media (tds).
- 3 Click Add.
- 4 In the Number of species text field, type 2.
- 5 Click 🔿 Study.
- 6 In the Select Study tree, select General Studies>Time Dependent.
- 7 Click 🗹 Done.

# ROOT

Add the model parameters from a text file.

## GLOBAL DEFINITIONS

Parameters 1

- I In the Model Builder window, under Global Definitions click Parameters I.
- 2 In the Settings window for Parameters, locate the Parameters section.
- 3 Click 📂 Load from File.
- **4** Browse to the model's Application Libraries folder and double-click the file liquid\_chromatography\_1\_parameters.txt.

The injection of the sample into the column is modeled with a Gaussian pulse.

#### Injection pulse

I In the Home toolbar, click f(X) Functions and choose Global>Gaussian Pulse.

- 2 In the Settings window for Gaussian Pulse, type Injection pulse in the Label text field.
- 3 In the Function name text field, type p1.
- 4 Locate the Parameters section. In the Location text field, type 3.

Variables I

- I In the Model Builder window, right-click Global Definitions and choose Variables.
- 2 In the Settings window for Variables, locate the Variables section.
- **3** In the table, enter the following settings:

Name	Expression	Unit	Description
pulse_inj	2.5*p1(t/1[s])		Time-dependent injection pulse with amplitude 1
rho_c	rho_p*(1-eps_p)	kg/m³	Density media in column

## GEOMETRY I

Interval I (i1)

- I In the Model Builder window, under Component I (compl) right-click Geometry I and choose Interval.
- 2 In the Settings window for Interval, locate the Interval section.
- 3 In the table, enter the following settings:

#### Coordinates (m)

0

- L\_C
- 4 Click 🟢 Build All Objects.
- **5** Click the **Com Extents** button in the **Graphics** toolbar.

## TRANSPORT OF DILUTED SPECIES IN POROUS MEDIA (TDS)

#### Porous Medium I

In the Model Builder window, under Component I (compl)>

Transport of Diluted Species in Porous Media (tds) click Porous Medium I.

## Adsorption I

- I In the Physics toolbar, click Attributes and choose Adsorption.
- 2 In the Settings window for Adsorption, locate the Adsorption section.
- **3** Select the **Species c1** check box.

- 4 Select the **Species c2** check box.
- 5 Locate the Matrix Properties section. Find the Density subsection. From the ρ list, choose User defined. In the associated text field, type rho\_c.
- 6 Locate the Adsorption section. In the  $K_{L,c1}$  text field, type K1.
- 7 In the  $c_{P,max,c1}$  text field, type S\*n01.
- **8** In the  $K_{L,c2}$  text field, type K2.
- **9** In the  $c_{P,max,c2}$  text field, type S\*n02.

# Fluid I

- I In the Model Builder window, expand the Porous Medium I node, then click Fluid I.
- 2 In the Settings window for Fluid, locate the Convection section.
- **3** Specify the **u** vector as

# v\_l x

- **4** Locate the **Diffusion** section. In the  $D_{\text{F,c1}}$  text field, type D\_1.
- **5** In the  $D_{\text{F,c2}}$  text field, type D\_2.

#### Porous Matrix I

- I In the Model Builder window, click Porous Matrix I.
- 2 In the Settings window for Porous Matrix, locate the Matrix Properties section.
- **3** From the  $\varepsilon_p$  list, choose **User defined**. In the associated text field, type eps\_p.

#### Inflow I

I In the Physics toolbar, click — Boundaries and choose Inflow.

The investigated sample is injected at the leftmost boundary.

- **2** Select Boundary 1 only.
- 3 In the Settings window for Inflow, locate the Concentration section.
- 4 In the c<sub>0.c1</sub> text field, type c01\*pulse\_inj.
- **5** In the  $c_{0,c2}$  text field, type c02\*pulse\_inj.

## Outflow I

- I In the Physics toolbar, click Boundaries and choose Outflow.
- **2** Select Boundary 2 only.

#### MESH I

Edge *I* In the **Mesh** toolbar, click **A** Edge.

#### Size

- I In the Model Builder window, click Size.
- 2 In the Settings window for Size, locate the Element Size section.
- 3 Click the **Custom** button.
- 4 Locate the Element Size Parameters section. In the Maximum element size text field, type 1e-4.
- 5 Click 📗 Build All.

# STUDY I

#### Step 1: Time Dependent

- I In the Model Builder window, under Study I click Step I: Time Dependent.
- 2 In the Settings window for Time Dependent, locate the Study Settings section.
- 3 In the **Output times** text field, type range(0,1,420).
- **4** From the **Tolerance** list, choose **User controlled**.
- 5 In the **Relative tolerance** text field, type 0.0010.

#### Solution 1 (soll)

- I In the Study toolbar, click The Show Default Solver.
- 2 In the Model Builder window, expand the Solution I (soll) node.
- 3 In the Model Builder window, expand the Study I>Solver Configurations> Solution I (soll)>Dependent Variables I node, then click Concentration (compl.cl).
- 4 In the Settings window for Field, locate the Scaling section.
- **5** From the **Method** list, choose **Manual**.
- 6 In the Scale text field, type c01.
- 7 In the Model Builder window, under Study I>Solver Configurations>Solution I (soll)> Dependent Variables I click Concentration (compl.c2).
- 8 In the Settings window for Field, locate the Scaling section.
- 9 From the Method list, choose Manual.
- **IO** In the **Scale** text field, type c02.

- II In the Model Builder window, under Study I>Solver Configurations>Solution I (soll) click Time-Dependent Solver I.
- **12** In the **Settings** window for **Time-Dependent Solver**, click to expand the **Absolute Tolerance** section.
- **I3** From the **Tolerance method** list, choose **Manual**.
- **I4** In the **Absolute tolerance** text field, type 1e-4.
- **I5** In the **Study** toolbar, click **= Compute**.

## RESULTS

Concentration, c1 (tds)

Follow these steps to reproduce the plot in Figure 3:

- I In the Model Builder window, under Results click Concentration, cl (tds).
- 2 In the Settings window for ID Plot Group, locate the Data section.
- 3 From the Time selection list, choose Interpolated.
- 4 In the Times (s) text field, type 5 50 100 200 300 385.
- 5 Click to expand the Title section. From the Title type list, choose None.
- 6 Locate the Plot Settings section. Select the x-axis label check box.
- 7 In the associated text field, type Column length (m).
- 8 Locate the Axis section. Select the Manual axis limits check box.
- **9** In the **y minimum** text field, type **0**.

#### Line Graph 1

- I In the Model Builder window, expand the Concentration, cl (tds) node, then click Line Graph I.
- 2 In the Settings window for Line Graph, click to expand the Coloring and Style section.
- 3 In the Width text field, type 2.
- 4 Click to expand the Legends section. Select the Show legends check box.
- 5 In the Concentration, cl (tds) toolbar, click **O** Plot.

Follow these steps to reproduce the plot in Figure 4:

## Concentrations, All Species (tds)

- I In the Model Builder window, under Results click Concentrations, All Species (tds).
- 2 In the Settings window for ID Plot Group, locate the Data section.
- 3 From the Time selection list, choose Interpolated.

- 4 In the Times (s) text field, type 10 100 200 300.
- 5 Locate the Title section. From the Title type list, choose None.

## Species c1

- I In the Model Builder window, expand the Concentrations, All Species (tds) node, then click Species c1.
- 2 In the Settings window for Line Graph, locate the Coloring and Style section.
- **3** In the **Width** text field, type **2**.
- 4 Locate the Legends section. Find the Include subsection. Select the Solution check box.
- **5** Clear the **Expression** check box.

#### Species c2

- I In the Model Builder window, click Species c2.
- 2 In the Settings window for Line Graph, locate the Coloring and Style section.
- **3** Find the Line style subsection. From the Line list, choose Dashed.
- **4** From the **Color** list, choose **Cycle (reset)**.
- **5** In the **Width** text field, type **2**.
- 6 Locate the Legends section. Clear the Show legends check box.

#### Concentrations, All Species (tds)

- I In the Model Builder window, click Concentrations, All Species (tds).
- 2 In the Settings window for ID Plot Group, locate the Plot Settings section.
- 3 Select the x-axis label check box.
- **4** In the associated text field, type Column length (m).
- 5 Locate the Axis section. Select the Manual axis limits check box.
- 6 In the **y minimum** text field, type 0.
- 7 In the Concentrations, All Species (tds) toolbar, click 💿 Plot.

Follow these steps to reproduce the plot in Figure 5:

## Detected concentration

- I In the Home toolbar, click 🚛 Add Plot Group and choose ID Plot Group.
- 2 In the Settings window for ID Plot Group, type Detected concentration in the Label text field.
- 3 Locate the Title section. From the Title type list, choose None.

## Point Graph 1

- I Right-click Detected concentration and choose Point Graph.
- **2** Select Boundary 2 only.
- 3 In the Settings window for Point Graph, locate the y-Axis Data section.
- **4** In the **Expression** text field, type c1+c2.
- 5 Click to expand the Coloring and Style section. In the Width text field, type 2.

#### Detected concentration

- I In the Model Builder window, click Detected concentration.
- 2 In the Settings window for ID Plot Group, locate the Plot Settings section.
- 3 Select the y-axis label check box.
- 4 In the associated text field, type Concentration (mol/m<sup>3</sup>).
- **5** In the **Detected concentration** toolbar, click **O Plot**.

## Concentrations, All Species (tds)

Last, an animation of the liquid chromatography process can be set up in the following manner:

#### Liquid chromatography animation

- I In the **Results** toolbar, click **IIII** Animation and choose File.
- **2** In the **Settings** window for **Animation**, type Liquid chromatography animation in the **Label** text field.
- 3 Locate the Scene section. From the Subject list, choose Concentrations, All Species (tds).
- 4 Locate the Target section. From the Target list, choose Player.
- 5 Locate the Animation Editing section. From the Time selection list, choose Interpolated.
- 6 Click Range.
- 7 In the Range dialog box, type 0 in the Start text field.
- 8 In the Step text field, type 2.
- **9** In the **Stop** text field, type **420**.
- IO Click Replace.
- **II** Click the **Play** button in the **Graphics** toolbar.

# 16 | LIQUID CHROMATOGRAPHY